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For research use only

# INSTRUCTION FOR USE REAL-TIME PCR DETECTION KITS

#### General information

#### Intended use:

The **Real-Time PCR Detection Kits** are *in vitro* DNA tests, which are intended for the specific identification of various microorganisms in human biological samples.

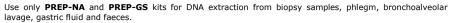
The kits are for research use only.

#### Method:

Real-time polymerase chain reaction; qualitative detection.

#### DNA extraction:

The "DNA-Technology" **PREP-NA**, **PREP-GS** and **PREP-RAPID** extraction kits are recommended. Some types of the samples must be pretreated (refer to the corresponding user manuals).



We do not recommend PREP-RAPID DNA Extraction Kit for DNA extraction from male urogenital swabs.

## Features:

PCR-Mix contains an internal control (IC). IC is intended for PCR quality and sufficiency of DNA assurance.

Positive control plasmid (C+) supplied with the kit is intended for specific PCR assessment.

We also recommend including in assay the negative control (C-) which is not supplied but very helpful for contamination control purposes. Use deionized water or sterile buffered saline instead of sample, starting from extraction step.

#### Devices:

The automatic analysis for **Real-Time PCR Detection Kits** is available on "DNA-Technology" made DTlite<sup>1</sup>, DTprime<sup>2</sup> Thermal Cyclers; the latest version of the software is available for download at <a href="https://www.dna-technology.com/software">https://www.dna-technology.com/software</a> or Bio-Rad Laboratories iCycler iQ thermal cyclers.

#### Time of analysis (excluding sample preparation procedure):

from 1.5 h.

#### The number of tests:

48/96 (including one positive control and one negative control in each run).

### Kits content

Reagent	Organoleptic parameters	48 tests Quantity 96 tests			
1. Paraffin sealed PCR-mix	Colorless transparent liquid under white wax layer	48 tubes or 6 8-tube-strips (20 μL in each tube)	96 tubes or 12 8-tube-strips (20 µL in each tube)		
2. Taq-polymerase solution	Colorless transparent liquid	1 tube (total volume 500 μL)	2 tubes (500 µL in each tube)		
3. Mineral oil	Colorless transparent viscous oily liquid	1 tube (total volume 1.0 mL)	2 tubes (1.0 mL in each tube)		
4. Positive control <sup>3</sup>	Colorless transparent liquid	1 tube (total volume 75 μL)	1 tube (total volume 150 μL)		
Associated accessories: Strip's caps <sup>4</sup>		6 8-caps	12 8-caps		

## Dye label detection channels

Fam	Hex	Rox	Cy5	Cy5.5
Specific product	IC	-	-	-

<sup>1 - 4</sup>S1, 4S2, 5S2, 6S1, 6S2 models.

<sup>&</sup>lt;sup>2</sup> - 4M1, 4M3, 4M6, 5M1, 5M3, 5M6, 6M1, 6M3, 6M6 models.

<sup>3 -</sup> marking as C+ is allowed.

<sup>&</sup>lt;sup>4</sup> - for detection kit packaged in strips.

#### Samples

REF	Microorganism	Sample		
R1-P002-23/4EU R1-P002-S3/4EU	Bordetella pertussis	Swabs from posterior pharyngeal wall		
R1-P406-23/4EU R1-P406-S3/4EU	Chlamydophila pneumoniae	Phlegm, bronchoalveolar lavage, swabs and washouts from nasal and oral pharynx		
R1-P401-23/4EU R1-P401-S3/4EU	Corynebacterium diphtheriae - toxigenic strains	Swabs from fibrin filmoral pharynx, larynx and other mucous membranes, lesion fluid		
R1-P322-23/9EU R1-P322-S3/9EU	Human papillomavirus 6 (HPV6)			
R1-P323-23/9EU R1-P323-S3/9EU	Human papillomavirus 11 (HPV11)	Swabs from urethra, cervix, nasal and oral pharynx, biopsy samples etc.		
R1-P301-23/9EU R1-P301-S3/9EU	Human papillomavirus 16 (HPV16) Human papillomavirus 18 (HPV18)			
R1-P011-23/4EU R1-P011-S3/4EU	Human Parvovirus B19	Blood plasma, saliva, biopsy samples from heart valves, amniotic fluid, autopsy samples		
R1-P003-23/4EU R1-P003-S3/4EU	Listeria monocytogenes	Spinal fluid, mucosal swabs, amniotic fluid, meconium, biopsy samples		
R1-P411-23/4EU R1-P411-S3/4EU	Mycoplasma pneumoniae	Phlegm, bronchoalveolar lavage, swabs and washouts from nasal and oropharynx		
R1-P412-23/4EU R1-P412-S3/4EU	Streptococcus pneumoniae	Phlegm, bronchoalveolar lavage, swabs and washouts from nasal and oral pharynx		
R1-P402-23/4EU R1-P402-S3/4EU	Streptococcus pyogenes	Swabs from urethra, cervix, posterolateral vaginal wall, lesions and mucosa of the amygdales, posterior pharyngeal wall		
R1-P001-23/9EU R1-P001-S3/9EU	Toxoplasma gondii	Spinal fluid, biopsy samples, etc		

## **Procedure**

### 1 PCR amplification



The reagents and tubes should be kept away from direct sun light.

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In case of using tubes in strips, strictly observe the completeness of the strips and caps to them. Do not use the caps to the strips of the other kits!

- 1.1 Mark tubes with paraffin sealed PCR-mix for test samples, negative control (C-) and positive control (C+).
  - **Example**. If you need to test 5 samples, mark 5 tubes for samples, 1 tube for "C-" and 1 tube for "C+". Total number of tubes 7.
- **1.2** Mix the Tag-polymerase solution thoroughly (3-5 s), then spin briefly (1-3 s).
- 1.3 Add 10 μL of Tag-polymerase solution into each tube. Avoid paraffin layer break.
- 1.4 Add one drop (~20 µL) of mineral oil into each tube. Close tubes.
- 1.5 Vortex the tubes with samples, "C-" and "C+" for 3-5 s and spin down drops for 1-3 s.



- 1. In case of using **PREP-GS DNA extraction kit**. After vortexing centrifuge the tubes with the DNA preparation at RCF(g) 16000 for one minute at room temperature (from 18  $^{\circ}$ C to 25  $^{\circ}$ C) to precipitate the sorbent. If, after isolation, the supernatant containing the isolated DNA was transferred to new tubes, centrifugation is carried out for 1-3 s on a vortex mixer.
- 2. Open the tube, add DNA sample (or control sample), then close the tube before proceeding to the next DNA sample to prevent contamination. In case of using tubes in strips, close the strip before proceeding to the next strips to prevent contamination. Use filter tips.
- 1.6 Add 5.0 μL of DNA sample into corresponding PCR-tubes. Avoid paraffin layer break. Use filter tips. Do not add DNA into the "C-". "C+" tubes.
- 1.7 Add  $5.0 \mu L$  of negative control (C-) which passed whole DNA extraction procedures and positive control (C+) into corresponding tubes. Avoid paraffin layer break.
- 1.8 Spin the tubes for 1-3 s.
- 1.9 Set the tubes to Real-time PCR Thermal Cycler.
- 1.10 For DTlite and DTprime thermal cyclers:

Launch the operating software for DT instrument<sup>5</sup>. Add corresponding test<sup>6</sup>, specify the number and ID's of the samples, positive and negative control samples. Specify the position of the tubes in the thermal unit (1.9) and run PCR (see table 1).

For iCycler iQ thermal cyclers:

Switch on the thermal and optical units of the device and let it warm up for 30 min. Launch iCycler (or Bio -Rad iQ5) software. Create and save new protocol if you do this protocol for the first time. In subsequent runs add the saved protocol, configure the plate (create the file with the data on samples ID's and position) and run PCR considering the total PCR reaction volume equal to 35 µL (see tables 2, 3).

<sup>&</sup>lt;sup>5</sup> Please, apply to Operation Manual for DTprime and DTlite Real-Time PCR instruments PART II.

<sup>&</sup>lt;sup>6</sup> Instructions for uploading "files with test parameters" can be found on "DNA-Technology's" website <a href="https://www.dna-technology.com/assaylibrary">https://www.dna-technology.com/assaylibrary</a>.

Table 1. The PCR program for DTlite and DTprime thermal cyclers

Step	Temperature, °C	Min.	Sec.	Number of cycles	Optical measurement	Type of the step
1	80.0 94.0	0	30 30	1		Cycle
2	94.0	0	30	5		Cycle
	64.0	0	15	3	V	Cycle
3	94.0	0	10	45		Cycle
J	64.0	0	15	7	V	Cycle
4	94.0	0	5	1		Cycle
5	10.0					Holding

Table 2. The PCR program for iCycler iQ5 thermal cyclers (with persistent well factor)

Cycle	Repeats	Step	Dwell time	Setpoint, °C	PCR/Melt Data Acquisition
1	1				
		1	1 min	80.0	
		2	1 min 30 sec	94.0	
2	5				
		1	30 sec	94.0	
		2	45 sec	64.0	
3	45				
		1	10 sec	94.0	
		2	45 sec	64.0	Real Time
4				10.0	Storage

Table 3. The PCR program for iCycler iQ thermal cyclers (with dynamic well factor)

Cycle	Repeats	Step	Dwell time	Setpoint, °C	PCR/Melt Data Acquisition
•		•	dynamicwf.tmo program		
1	1				
		1	1 min	80.0	
		2	1 min 30 sec	94.0	
2	5				
		1	30 sec	94.0	
		2	45 sec	64.0	
3	2				
		1	30 sec	80.0	Real Time
•		•	PCR program		
4	45				
		1	10 sec	94.0	
		2	45 sec	64.0	Real Time
5				10.0	Storage

# 2 Data collection and data analysis

Registration and interpretation of the PCR results is held in automatic mode.

# Storage, shipping and handling requirements



All components of the kits should be stored at the temperatures from 2 °C to 8 °C during the storage period.

Paraffin sealed PCR-mix must be stored at temperatures from 2  $^{\circ}\text{C}$  to 8  $^{\circ}\text{C}$  and out of light during the storage period.

Excessive temperature and light can be detrimental to product performance.

Transportation of the kit is allowed in termobox with ice packs by all types of roofed transport at temperatures from 2 °C to 25 °C but no more than 5 days and should be stored at temperatures from 2 °C to 8 °C immediately on receipt.

Shelf life - 12 months since the date of production in compliance with all transportation, storage and operation conditions.

Contact our customer service department regarding quality issues with the kit:

8 (800) 200.75.15 (toll-free call for Russia)

+7 (495) 640.16.93 (chargeable call for CIS and foreign countries).

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Address: 117587, Russia, Moscow, int. ter. Municipal District Chertanovo Severnoye, Varshavskoye shosse,

125 Zh, building 5, floor 1, office 12

#### Kev to symbols

RUO	For research use only	[]i	Consult instructions for use	REF	Catalogue number
1	Temperature limit	3	Manufacturer	LOT	Batch code
$\subseteq$	Use-by date	Σ	Contains sufficient for <n> tests</n>	**	Keep away from sunlight
$\sim$	Date of manufacture	NON	Non-sterile	$\triangle$	Caution

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